

(19)

Europäisches Patentamt
European Patent Office
Office européen des brevets



(11) EP 1 103 270 A1

(12)

EUROPEAN PATENT APPLICATION
published in accordance with Art. 158(3) EPC

(43) Date of publication:

30.05.2001 Bulletin 2001/22

(51) Int. Cl.: A61K 38/22, A61P 37/02

(21) Application number: 00931296.8

(86) International application number:
PCT/ES00/00197

(22) Date of filing: 02.06.2000

(87) International publication number:
WO 00/74708 (14.12.2000 Gazette 2000/50)

(84) Designated Contracting States:

AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU
MC NL PT SE

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Designated Extension States:

AL LT LV MK RO SI

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(30) Priority: 04.06.1999 ES 9901235

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(54) METHOD FOR TREATING ENDOTOXIC SHOCK AND INFLAMMATORY AND AUTOIMMUNE DISEASES IN MAMMALS

(57) The composition and use of therapeutic agents that inhibit the production of tumoral necrosis factor and interleukin 6 are described. Said agents induce high levels of interleukin 4 for the treatment of endotoxic shock and inflammatory and autoimmune diseases, respectively, in mammals. The composition includes substances such as vasoactive intestinal peptide, adenylate cyclase hypophysary activator peptide and fragments and derivatives thereof.

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Description**STATE OF THE ART**

[0001] Endotoxic shock is still the main cause of death in hospitals. The strategies for combating the effects of endotoxic shock focus on counteracting the bacterial agents responsible for the effect, on restoring the haemodynamic parameters, preventing cellular activation and on modifying the action of the defence mechanisms (Boyd O; Current Opinion in Anaesthesiology 1996, 9:98).

[0002] It is currently accepted that the inflammatory response to bacterial products directly contributes to the development of endotoxic shock (Parillo JE; New England Journal of Medicine 1993, 328:1471). The toxic bacterial products and those released during tissue damage activate the defence mechanisms, implicating cells such as neutrophils, monocytes, macrophages and endothelial cells, and mediators such as cytokines, platelet activation factor, metabolites of arachidonic acid and nitric oxide, causing haemodynamic changes and organics harmful to the host (Moldawer LL; Critical Care Medicine 1994, 22:3). Many cytokines have been proposed as markers of the seriousness of the development of septic shock. The circulating levels of TNF α , IL-1, IL-6 and IL-8 have been correlated with the probability of overcoming a septic episode. TNF α and IL-1 administered to humans or to experimental animals reproduce many of the haemodynamic manifestations of septic shock (Tracey KJ and co-workers; Science 1986, 234:470) and its inhibition has been assayed by means of injection of antagonist receptors and neutralising monoclonal antibodies with different results (Fisher CJ and co-workers; Critical Care Medicine 1994, 22:12). Among the immunological markers, the levels of circulating IL-6 are the best indicators of the seriousness of the sepsis and of the possibilities of recovering from the episode (Liaw YS and co-workers; Journal of the Formosan Medical Association 1997, 96:685).

[0003] Despite the advance in the knowledge of the mechanisms and of the technological and pharmacological progress there are still few results concerning an improvement in the data for death, which translate into a figure of around 200,000 per year in the United States and Europe (Vicent J-L and Chamlou R; Current Opinion in Anaesthesiology 1996, 9:146).

[0004] Inflammatory processes are a vital process for the survival of all complex organisms. Inflammation is a natural defence process of the organism against foreign agents. The accumulation and activation of leukocytes in places where the aggression takes place is a central occurrence in all inflammatory processes (Schaal TJ and Bacon KB; Current Opinion in Immunology 1994, 6:865). An insufficient inflammatory response may compromise the survival of the organism, but an excessive response, which may be due to failures in the

mechanisms of deactivation of the process due to different causes, can lead to an inflammatory or autoimmune disease (Saccà R and co-workers; Current Opinion in Immunology 1997, 9:851). These diseases are an important cause of morbidity and mortality in mammals due to tissue damage associated with said processes.

[0005] Macrophages play a key role in the regulation of immune and inflammatory responses. The execution of these activities is mediated by a whole series of complex processes in which, among others, many products of macrophage origin intervene. As a response to the antigens, and according to their origin, the macrophages secrete pro-inflammatory cytokines and oxidising agents, such as TNF α , IL-6, IL-1 β , IL-12 and nitric oxide (Laskin DL and co-workers; Annual Review of Pharmacology and Toxicology 1995, 35:655). TNF α and IL-6 are, among others, two factors that contribute to physiopathological changes associated with several states of chronic or acute inflammation. The macrophages, in addition, participate in the initiation, maintenance and control of immune responses, acting as potent antigen presenters, providing the T-lymphocytes with a double activation signal: the antigen-molecule complex of the main histocompatibility complex (MHC) and a co-stimulatory signal mediated by molecules of the B7 family (Lenschow DJ and co-workers; Annual Review of Immunology 1996, 14:233). The B7 molecules comprise two isoforms, B7.1 and B7.2, both of which are implicated in the stimulation of two different types of T helper cells (Th), Th1 and Th2, and both of these produce a set of different cytokines (Kuchroo VK and co-workers; Cell 1995, 80:707).

[0006] Activation of Th1 cells implies the production of IFN γ and IL-12, among other factors, and is associated with the production of antibodies of the IgG2a isotype and it is manifest as a reaction of delayed inflammatory type. The activation of Th2 cells implies the production of IL-4, IL-5 and IL-10, among other factors, is associated with the secretion of antibodies of the isotype IgG1, inhibits the delayed inflammatory response and is manifest as a humoral response (Constant SL y Bottomly K; Annual Review of Immunology 1997, 15:297). The factors that determine the differentiation of one or another type of response are mainly the characteristics of the antigen presenting cells and the cytokines present in the microenvironment in which the response takes place: IL-12 determines the differentiation of Th1 cells while IL-4 does so with Th2. When both are present, the IL-4 effect predominates (O'Garra AO; Immunity 1998, 8:275). Numerous cases of inflammatory and autoimmune diseases are due to the activation of an inappropriate type of Th cell.

[0007] The vasoactive intestinal peptide (VIP) is a basic peptide containing 28 amino acids units whose sequence is (Mutt V and Said SI; European Biochemistry 1974, 42:581):

His-Ser-Asp-Ala-Val-Phe-Thr-Asp-Asn-Tyr-Thr-

Arg-Leu-Arg-Lys-Gln-Met-Ala-Val-Lys-Lys-Tyr-Leu-Asn-Ser-Ile-Leu-Asn-NH₂

Ala-Val-Leu-Gly-Lys-Arg-Tyr-Lys-Gln-Arg-Val-Lys-Asn-Lys-NH₂

[0008] It was isolated firstly from the small intestine of pig and later was identified in the brain and in the nerve endings of the peripheral system. It was established that it was a neuropeptide with neuromodulating properties (Fahrenkrug J; Pharmacology and Toxicology 1993, 72:354). It takes its name from its peripheral vasodilatory properties. VIP has also been identified in mast cells of rat and in granulomas (Cutz E. and co-workers; Nature 1978, 275:661). Immunochemical studies carried out in histological sections of the thymus, spleen and lymphatic ganglia of rat have identified immunoreactive VIP in lymphocytes of these organs (Gomariz RP and co-workers; Annals of the New York Academy of Sciences 1992, 650:13; Leceta and co-workers; Advances in Neuroimmunology 1996, 6:29).

[0009] VIP exercises its biological effects by means of membrane receptors belonging to the superfamily of seven hydrophobic domains coupled to G proteins, which transduce the information to the final effector molecules (Laburthe M and Couvineau A; Annals of the New York Academy of Sciences 1988, 527:296). The receptors for VIP have been characterised in numerous tissues such as liver and adipose tissue, among others, and correspond to two types, the so-called VIP1-R (Ishihara T and co-workers; Neuron 1992, 8:811) and VIP2-R (Lutz E. and co-workers; FEBS Letters 1993, 334:3). In the immune system, receptors specific to VIP have been characterised in a variety of immune cells which include human peripheral lymphocytes, human monocytes, rat and mouse lymphocytes, alveolar macrophages of rat and peritoneal macrophages of rat and mouse (Gomariz RP and co-workers; Biochemical and Biophysical Research Communications 1994, 203:1599; Delgado M and co-workers; Regulatory Peptides 1996, 62:161). VIP modulates a wide variety of immune functions such as the phagocyte function, in each one of the stages of the process, the proliferate response, the production of immunoglobins, the NK activity and the production of cytokines (De La Fuente M and co-workers; Advances in Neuroimmunology 1996, 6:75).

[0010] The adenylate cyclase hypophysary peptide activator (ACHPA) is a member of the family of peptides of the secretin/VIP/glucagons of which two molecular forms are known: ACHPA-38 and ACHPA-27, whose sequences are respectively (Ogi K and co-workers; Biochemical and Biophysical Research communication 1993, 196:1511):

ACHPA-38

[0011]

His-Ser-Asp-Gly-Ile-Phe-Thr-Asp-Ser-Tyr-Ser-Arg-Tyr-Arg-Lys-Gln-Met-Ala-Val-Lys-Lys-Thy-Leu-Ala-

ACHPA-27

[0012]

His-Ser-Asp-Gly-Ile-Phe-Thr-Asp-Ser-Tyr-Ser-Arg-Tyr-Arg-Lys-Gln-Met-Ala-Val-Lys-Lys-Thy-Leu-Ala-Ala-Val-Leu-NH₂

[0013] Both peptides are widely distributed in the central nervous system and peripheral system.

[0014] There are also cells that produce ACHPA in the lung, pancreatic B cells and intestine (Arimura A; Regulatory Peptides 1992, 37:287). In the immune system, a large abundance of ACHPA positive cells has been described in central and peripheral lymphoid organs (Gaytan F and co-workers; Cell and Tissue Research 1994, 276:233). For ACHPA, three types of receptor have been described (Shivers BD and co-workers; Endocrinology 991, 128:3055; Inagaki N and co-workers; Proceeding of the National Academy of Sciences USA 1994, 91:2679). The type-1 ACHPA (ACHPA-R-I) with equal affinity for ACHPA-38 and ACHPA-27, but which has an affinity of 300 to 1000 times less for VIP; the type-2 ACHPA receptor (ACHPA-R-II) which recognises VIP, ACHPA-38 and ACHPA-27 with the same affinity, and so is denominated the VIP-ACHPA common receptor and corresponds to the receptor of VIP VIP1-R, and the type-III ACHPA receptor (ACHPA-R-III) which corresponds to the receptor of VIP VIP2-R. Until present, there are few studies on the biological actions of ACHPA in the immune system. The effects of ACHPA are often similar to those of VIP modulating the phagocyte function and the proliferate responses.

DESCRIPTION OF THE INVENTION

[0015] The object of this invention is to develop preparations of VIP, ACHPA and analogues as therapeutic agents in the treatment of endotoxic shock and inflammatory and autoimmune diseases.

[0016] The treatment consists of the administration to mammals, in need thereof, of an effective quantity of an agent that inhibits the production of tumoral necrosis factor (TNF) or IL-6 in a pharmaceutically acceptable vehicle, or else the administration to mammals, in need thereof, of an effective quantity of an agent that increases the production of IL-4, inhibiting the activation of Th1 cells and stimulating the activation of Th2 cells.

[0017] It is known that most of the effects of endotoxic shock are mediated by the activation of the immune system and the inflammatory mechanisms of the host as a response to the bacterial products. The macrophages play a very relevant role in this process as, after their activation, they produce factors such as

nitric oxide, prostaglandins and cytokines which are responsible for symptoms such as fever, hypotension, disseminated microcoagulation, multiple organ failure and finally death. In this sense, high circulating levels of TNF, IL-1 and IL-6 associated with endotoxaemia have been described. In animal models these symptoms are reproduced both by the administration of bacterial endotoxins (LPS) and by injection of TNF and IL-1. Other studies have shown the diagnostic value in terms of probability of survival that circulating levels of IL-6 represent (Stoiser B and co-workers; European Journal of Clinical Investigation 1998, 28:672).

[0018] The tumoral necrosis factor (TNF) is produced by several types of cell which include monocytes and macrophages, T and B lymphocytes, neutrophils, mast cells, tumoral cells and fibroblasts. It is an important regulatory factor of other pro-inflammatory cytokines, such as IL-1 β , IL-6 and IL-8. TNF α induces the expression of adhesion molecules in endothelial cells, activates the leukocytes so that they destroy the microorganisms, acts on the hepatocytes in order to increase synthesis of serum proteins which contribute to the response in acute phase and activate the coagulation system. Overproduction of this molecule leads to immunopathological diseases, autoimmunity and inflammation.

[0019] IL-6 is a multifunctional cytokine produced both by lymphocytes and by non-lymphoid cells. It regulates several aspects of the immune response, such as production of proteins that mediate the acute phase of haematopoiesis. In addition, it acts as a mediator in the inflammatory response. Its production is regulated by several factors which include TNF α , IL-1 and bacterial endotoxin (LPS).

[0020] IL-4 is a cytokine that inhibits the production of pro-inflammatory cytokines, promotes the proliferation and differentiation of activated lymphocyte B molecules and increases the expression of MHC molecules of type II and B lymphocytes. Its possible clinical use in anti-inflammatory treatment and autoimmune diseases has been highlighted.

[0021] Strategies have been tried for neutralising pro-inflammatory cytokines in the treatment of endotoxic shock but the results do not indicate that there is greater survival in the long term. A treatment that inhibits the production of TNF α and IL-6 would represent a considerable improvement in the evolution of endotoxic shock and in the probabilities of survival. The administration of VIP and ACHPA in animal models achieved these effects and our invention consists in the use of a treatment with these neuropeptides to increase the survival in endotoxic shock and revert pathological inflammatory states and autoimmune diseases.

VIP and ACHPA have anti-inflammatory effects and inhibit the production of IL-6 and TNF α in animal models of induction of endotoxic shock. As these cytokines play an important role in the development of said syndrome, VIP and ACHPA can be used to regulate their

5 production. In addition, VIP and ACHPA modulate the capacity of the antigen presenting cells to act inducing the activation of proliferation and differentiation of lymphocytes with a pattern of cytokine secretion typical of Th2 cells and condition the immune responses "in vivo" favouring the development of response of the humoral type.

DESCRIPTION OF THE FIGURES

10 [0022]

Figure 1 represents the production of TNF α by murine macrophages in culture (5×10^5 cells/ml) stimulated with 10ng/ml of LPS in the presence or absence of 10^{-8} M of VIP or ACHPA during the course of 24 hours.

15 Figure 2 represents the production of TNF α by murine macrophages in culture (5×10^5 cells/ml) after 6 hours of culture with 10ng/ml of LPS and to which 10^{-8} M of VIP or ACHPA are added at different times.

20 Figure 3 represents the production of IL-6 by murine macrophages in culture (5×10^5 cells/ml) stimulated with 10ng/ml of LPS in the presence or absence of 10^{-8} M of VIP or ACHPA during the course of 24 hours.

25 Figure 4 represents the production of IL-6 by murine macrophages in culture (5×10^5 cells/ml) after 6 hours of culture with 10ng/ml of LPS and to which 10^{-8} M of VIP or ACHPA are added at different times.

30 Figure 5 presents the Northern Blot analysis for the presence of mRNA of TNF α and IL-6 in macrophages stimulated with LPS in the presence or absence of VIP or ACHPA (18S represents the corresponding rRNA as total quality control of loaded RNA).

35 Figure 6 represents the survival in mice injected with 400 μ g of LPS and, simultaneously, or after 30 minutes, 1 or 4 hours, with 5 nmol of VIP or ACHPA. A: Control; B: VIP at 0 h.; C: VIP at 0,5 h; D: VIP at 1 h.; E: VIP at 4 h.

40 Figure 7 represents the number of IL-4 secreting cells in the spleen and peritoneum detected by means of the immunabsorption assay technique in conjugated plate with enzymes (ELISPOT) in mice immunised in the conditions specified in Example 7 and which simultaneously to the second injection of antigen received 5 nmol of VIP or ACHPA or an injection of saline solution.

Figure 8 represents the quantity of anti-haemocyanine immunoglobulins of snail (anti-KLH) of the iso-types IgG2a and IgG1 detectable in serum by means of the technique conjugated immunoabsorption assays with enzymes (ELISA) in immunised mice in the conditions specified in Example 8 and with the serum samples taken two weeks after the last injection.

Figure 9 represents the number of IL-4 producing cells detected by means of the ELISPOT technique in mice that were immunised in the conditions specified in Examples 7 and 8 and which, in the second injection, received or did not receive 5 nmol of VIP along with 100 μ gr of IgG, anti-B7.1 or anti-B7.2.

EMBODIMENT OF THE INVENTION

[0023] The following examples are only to illustrate the results obtained and do not limit the use of the invention which is detailed in the specified claims.

EXAMPLE 1

VIP and ACHPA inhibit the production of TNF α in macrophages stimulated with LPS

[0024] In experiments carried out "in vitro" VIP and ACHPA inhibit the production of TNF α in peritoneal murine macrophages stimulated with LPS. The greatest degree of inhibition reaches values near to 60% and occurs with doses of stimulation between 1-10 ngr./ml of LPS. The IC₅₀ value is 80 pM, both for VIP and for ACHPA and its effect was observed up until the end of the experiment (see Figure 1). The inhibitory effect is the same if both neuropeptides are added up until 1 hour after stimulating the macrophages with LPS, although it progressively reduces until disappearing if they are added after 4 hours (see Figure 2).

EXAMPLE 2

VIP and ACHPA reduce the circulating levels of TNF α after injection of LPS

[0025] In an experiment carried out with mice the circulating levels of TNF α 2 hours after injection of 25 μ gr of LPS were approximately 4 ngr./ml. Simultaneous administration of 5 nmol of VIP or ACHPA reduced said levels by 60%.

EXAMPLE 3

VIP and ACHPA inhibit production of IL-6 in macrophages stimulated with LPS

[0026] In experiments carried out "in vitro" VIP and

5 ACHPA inhibit the production of IL-6 in peritoneal murine macrophages stimulated with LPS. Most of the inhibition reaches values near to 90% and occurs with doses of stimulation of 10 μ gr/ml of LPS. The IC₅₀ is 8.6 pM, both for VIP and for ACHPA and the effect was observed up until the end of the experiment (see Figure 3). The inhibitory effect is also observed if the neuropeptides are added after stimulation with LPS, although the degree of inhibition is progressively smaller (see Figure 4).

EXAMPLE 4

VIP and ACHPA reduce the circulating levels of IL-6 after injection of LPS

[0027] 20 In an experiment carried out in mice the circulating levels of IL-6 two hours after injection of 25 μ gr. of LPS were approximately 1.5 ngr./ml. The simultaneous administration of 5nmol of VIP or ACHPA reduced said levels by 60% and 75%, respectively.

EXAMPLE 5

VIP and ACHPA regulate the production of TNF α and IL-6 at a transcriptional level

[0028] 30 Macrophages from rat were submitted to the experimental conditions described in examples 1 and 3 and their mRNA was isolated. This mRNA was analysed using the Northern Blot technique to detect mRNA of TNF α and IL-6. Figure 5 shows the absence of transcripts for TNF α or IL-6 when the macrophages activated with LPS are also exposed to VIP or ACHPA.

EXAMPLE 6

VIP and ACHPA protect against the lethal effects of LPS

[0029] 40 An experiment was carried out in which the long-term survival over a period of 4 days was studied for mice injected with 400 μ gr of LPS. The results are reflected in figure 6.

[0030] 45 The mortality in these circumstances was 100% after 36 hours. With the simultaneous administration of 5 nmol of VIP or ACHPA a survival of 60% was achieved at the end of the experiment. The administration of neuropeptides up to 1 hour after injection with LPS still gave survival rates close to 50%.

EXAMPLE 7

VIP and ACHPA increase the proportion of IL-4 secreting cells.

[0031] 55 Groups of mice were immunised with 50 μ gr of KLH emulsified in an adjuvant, repeating the injection with 100 μ gr of KLH two weeks later and simultaneously

injecting 5 nmol/mouse of VIP, ACHPA or saline solution. Two weeks after the last injection, suspensions were made of the spleen and peritoneum cells. These were cultured for 24 hours in the presence of 50 µgr/ml of KLH, after which time the number of IL-4 producing cells was determined using the ELISPOT technique. In the mice injected with VIP or ACHPA, the number of IL-4 producing cells increased in the order of 20 times compared to those that were not treated with these neuropeptides (see Figure 7).

EXAMPLE 8

VIP and ACHPA induce the production of antibodies of the isotype IgG1.

[0032] Groups of mice were immunised with 50 µgr of KLH emulsified in an adjuvant, repeating the injection with 100 µgr of KLH two weeks later and simultaneously injecting 5 nmol/mouse of VIP, ACHPA or saline solution. Two weeks after the last injection, the levels of anti-KLH and its isotype were determined using ELISA specific to the IgG1 and IgG2a isotypes. In mice injected with VIP or ACHPA the anti-KLH antibodies detectable in serum two weeks after the last immunisation are only of the IgG1 isotype, while in those that only received saline solution they were of the isotype IgG2a (see Figure 8)

EXAMPLE 9

The increase in the proportion of IL-4 producing cells mediated by VIP and ACHPA is related to the expression of B7.2 induced by both neuropeptides.

[0033] Groups of mice were immunised in the same conditions as those for Examples 7 and 8, but in the moment of the second immunisation with KLH the mice that were simultaneously injected with VIP or ACHPA also received 100 µgr of anti-B7.1, anti-B7.2 antibody or the same quantity of IgG as control. In the mice that received anti-B7.2 antibodies simultaneously to the administration of neuropeptides the number of IL-4 producing cells was reduced to the proportion reached in animals that were not injected with neuropeptides (see Figure 9).

Claims

1. A method for the treatment of endotoxic shock in mammals characterised in that it comprises the administration of an effective quantity of an agent that inhibits the production of tumoral necrosis factor (TNF) in a pharmaceutically acceptable vehicle.
2. A method for the treatment of endotoxic shock in mammals according to claim 1, characterised in that the inhibitory agent is a vasoactive intestinal

peptide (VIP) or any fragments thereof or some analogue derivative.

3. A method for the treatment of endotoxic shock in mammals according to claim 1, characterised in that the inhibitory agent is the adenylate cyclase hypophysary peptide activator (ACHPA) or any fragments thereof or some analogue derivative.
4. A method for the treatment of endotoxic shock in mammals characterised in that it comprises the administration of an effective quantity of an agent that inhibits the production of interleukin 6 (IL-6) in a pharmaceutically acceptable vehicle.
5. A method for the treatment of endotoxic shock in mammals according to claim 4, characterised in that the inhibitory agent is a vasoactive intestinal peptide (VIP) or any fragments thereof or some analogue derivative.
6. A method for the treatment of endotoxic shock in mammals according to claim 4, characterised in that the inhibitory agent is the adenylate cyclase hypophysary peptide activator (ACHPA) or any fragments thereof or some analogue derivative.
7. A method for the treatment of inflammatory or autoimmune pathologies in mammals, characterised by the activation of Th1 cells, which comprises the administration of an effective dose of an agent, in a pharmaceutically appropriate vehicle, which induces high levels of IL-4.
8. A method for the treatment of inflammatory or autoimmune pathologies in mammals according to claim 7, characterised in that the inducing agent is the vasoactive intestinal peptide (VIP) or a fragment thereof or some analogue derivative.
9. A method for the treatment of inflammatory or autoimmune pathologies in mammals according to claim 7, characterised in that the inducing agent is the adenylate cyclase hypophysary activator peptide (ACHPA) or any fragment thereof or some analogue derivative.

Amended claims under Art. 19.1 PCT

- 50 1. Use of the vasoactive intestinal peptide (VIP) or any of its fragments or some analogue derivative for the preparation of a drug destined for the treatment of endotoxic shock in mammals, due to their capacity as agents that inhibit the production of the tumoral necrosis factor (TNF) and interleukin 6 (IL-6).
- 55 2. Use of the adenylate cyclase hypophysary acti-

vator peptide (ACHPA) or a fragment thereof or a derivative for the preparation of a drug destined to the treatment of endotoxic shock in mammals, due to their capacity as agents that inhibit the production of the tumoral necrosis factor (TNF) and interleukin 6 (IL-6). 5

3. Use of the vasoactive intestinal peptide (VIP) or any of its fragments or some analogue derivative for the preparation of a drug destined to the treatment of inflammatory or autoimmune pathologies characterised by the activation of Th1 cells, such as rheumatoid arthritis, multiple sclerosis, Crohn's disease, implant reaction to host and others, due to their capacity as inhibitors of Th1 cells. 10

4. Use of the adenylate cyclase hypophysary activator peptide (ACHPA) or a fragment thereof or a derivative for the preparation of a drug destined to the treatment of inflammatory or autoimmune pathologies characterised by the activation of Th1 cells, such as rheumatoid arthritis, multiple sclerosis, Crohn's disease, implant reaction to host and others, due to their capacity as inhibitors of Th1 cells. 20

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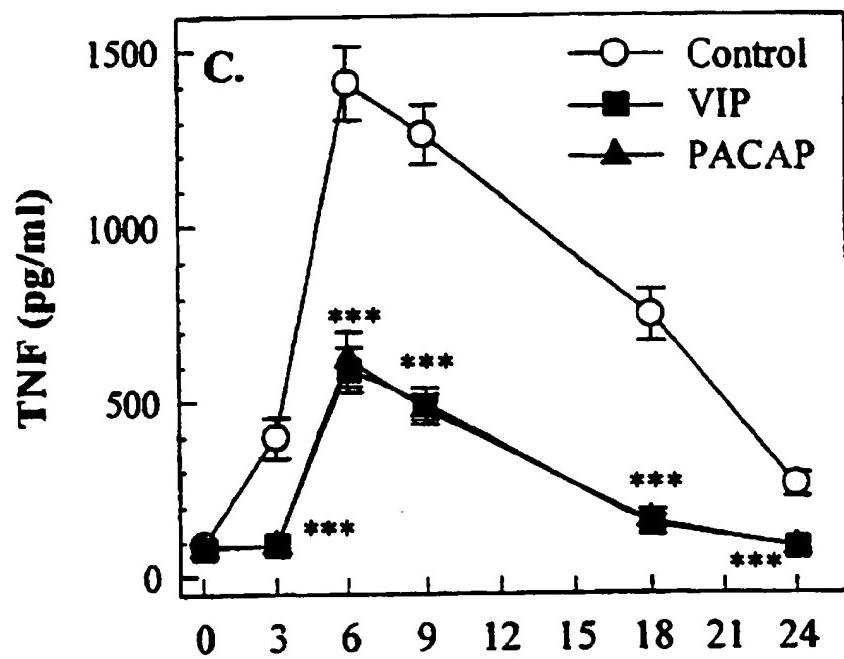


FIGURA 1

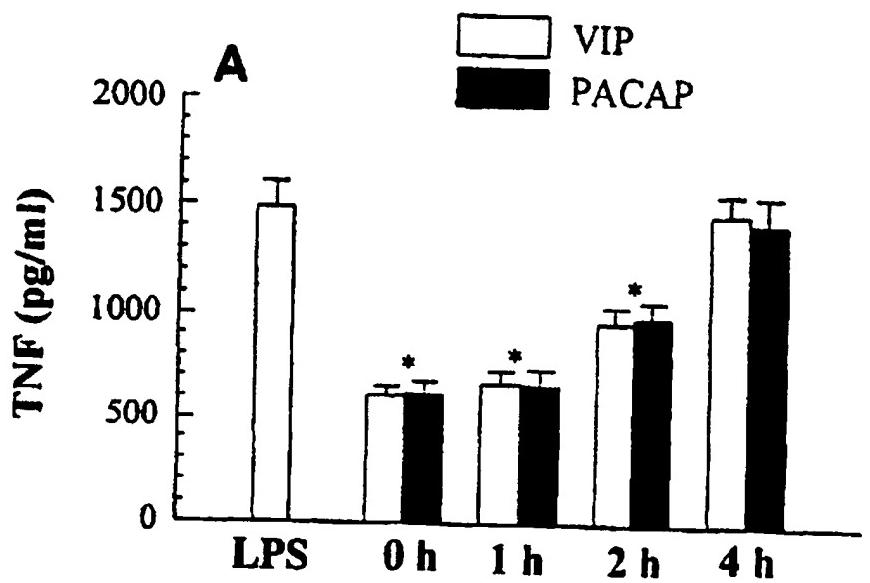


FIGURA 2

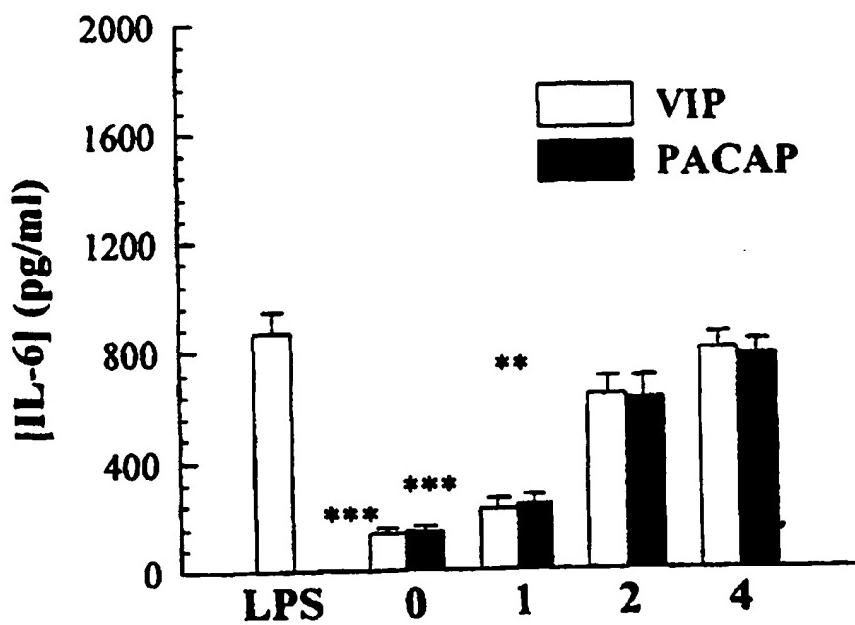


FIGURA 3

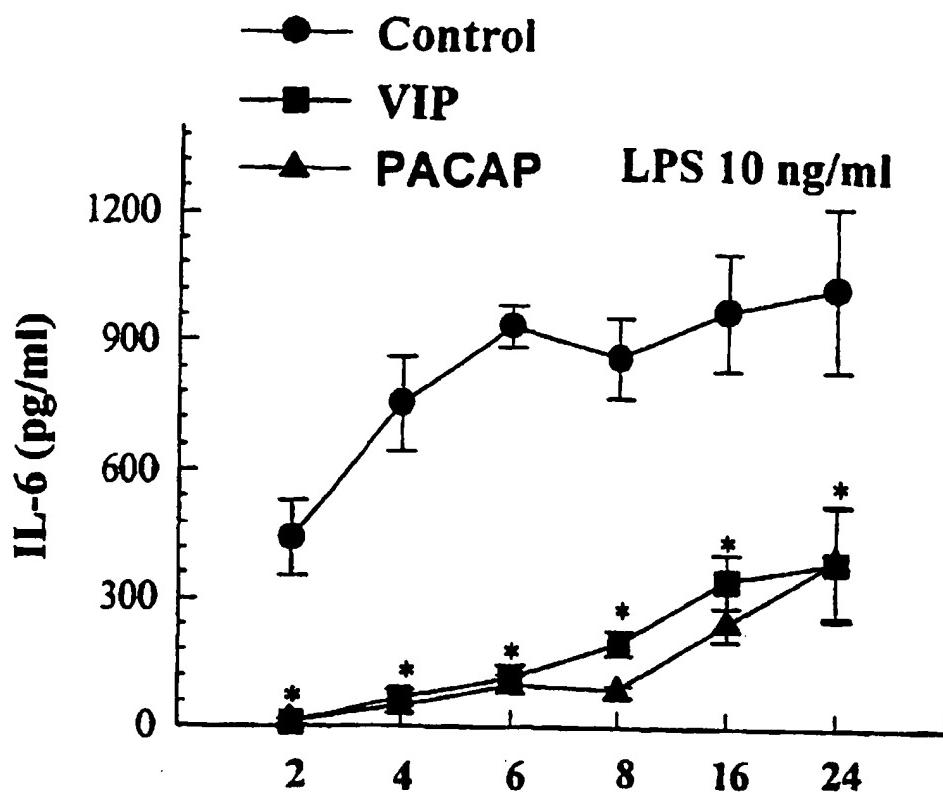


FIGURA 4

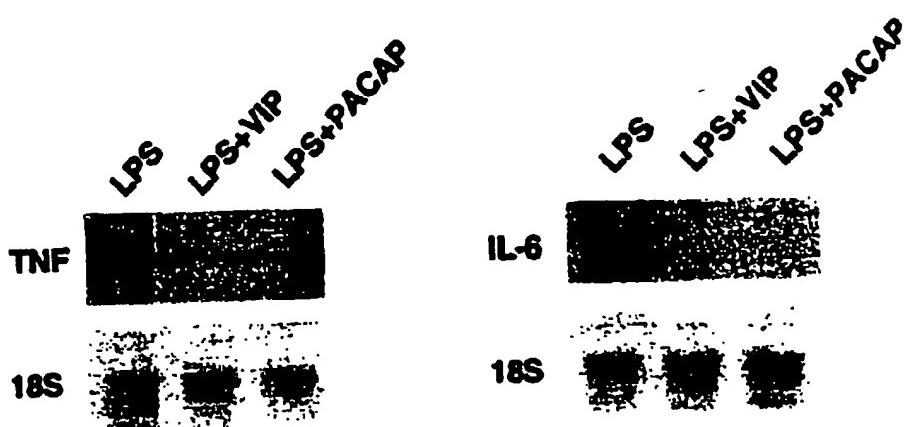


FIGURA 5

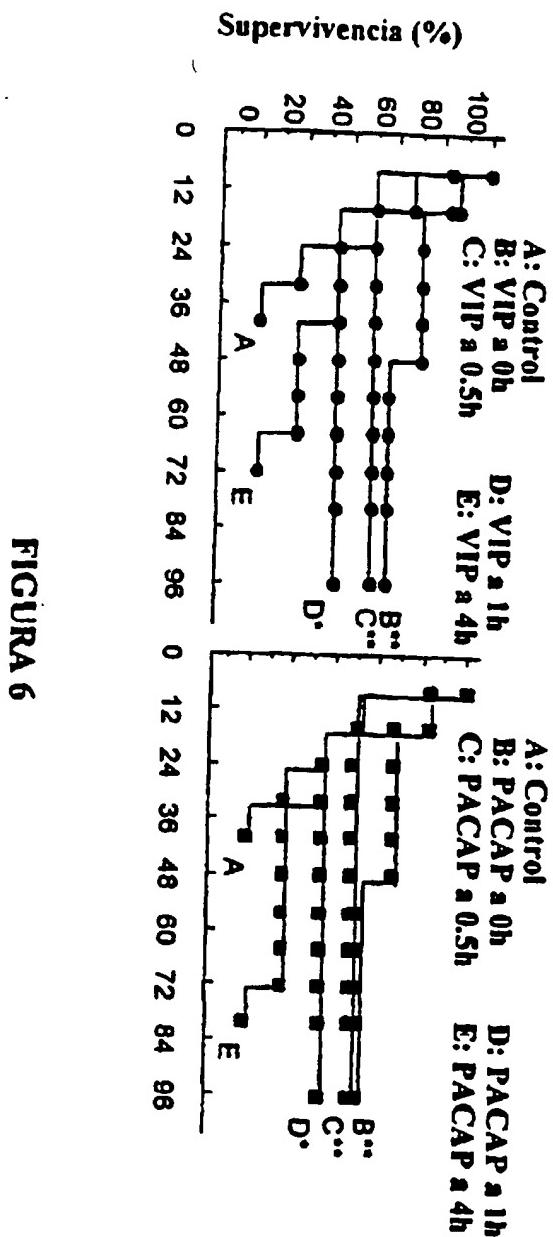


FIGURA 6

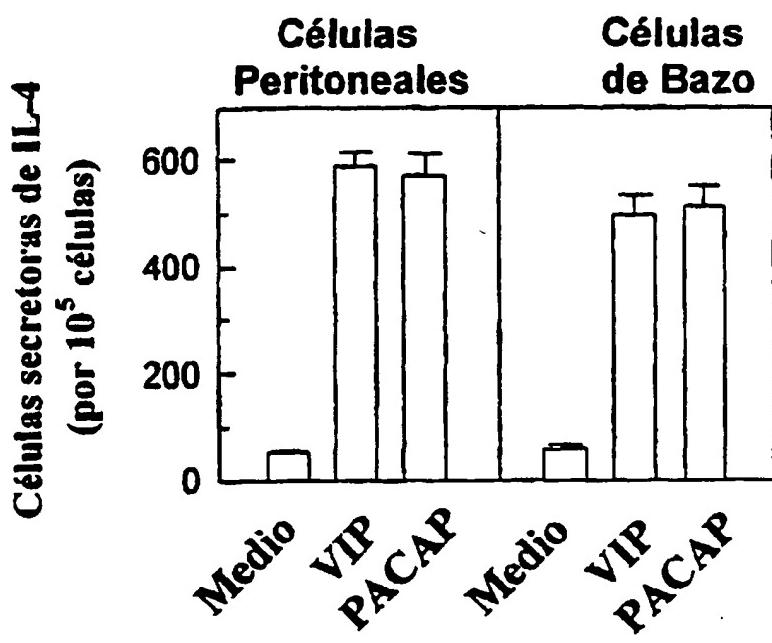


FIGURA 7

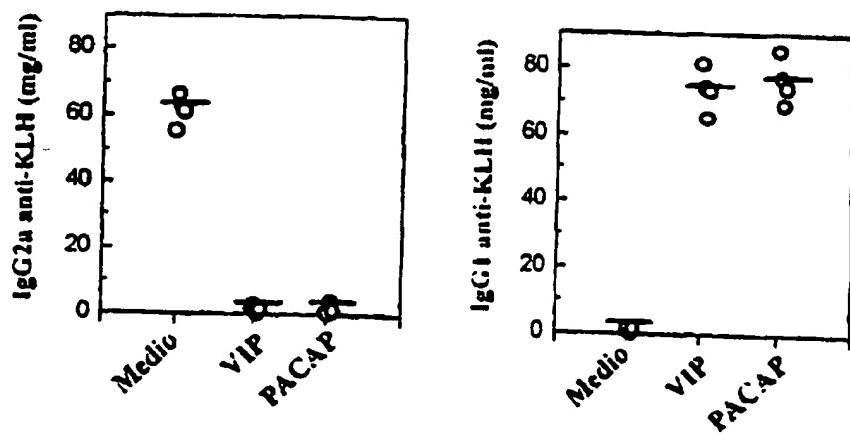


FIGURA 8

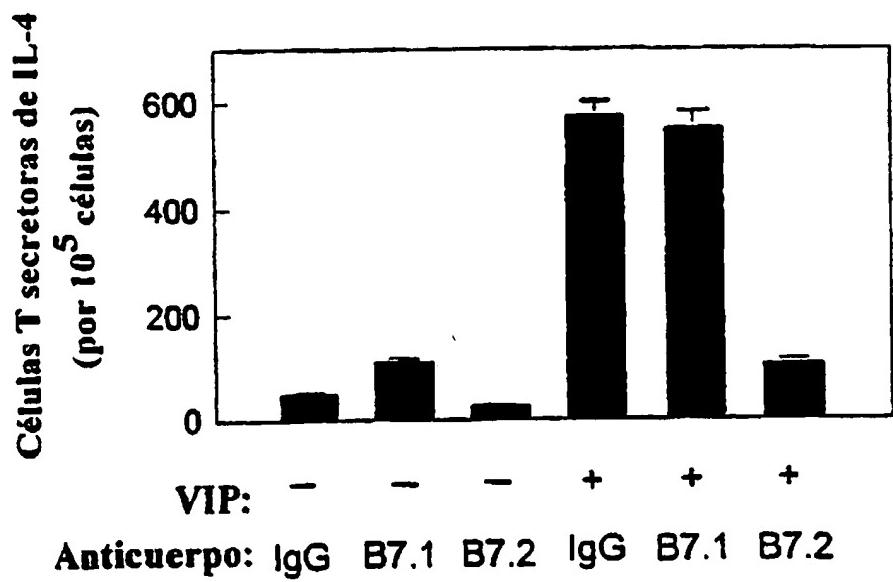


FIGURA 9

INTERNATIONAL SEARCH REPORT

International application No.
PCT/ES 00/00197

A. CLASSIFICATION OF SUBJECT MATTER IPC 7 A61K 38/22, A61P 37/02 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 7 A61K, A61P, C07K Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) EPODOC, WPIL, CA, BIOSIS, EMBASE, MEDLINE		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
E	WO 9953944 A1 (Universidad Complutense de Madrid Rectorado), 28 October 1999 (28.10.1999), the whole document, specially claims 1-6	1-6
X	GANEA, D. "Regulatory effects of vasoactive intestinal peptide on cytokine production in central and peripheral lymphoid organs". ADVANCES IN NEUROIMMUNOLOGY. 1996, Vol. 6, n 1, pages 61-74. The whole document	7-9
<input type="checkbox"/> Further documents are listed in the continuation of box C. <input checked="" type="checkbox"/> Patent family members are listed in annex.		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed		
Date of the actual completion of the international search 24 August 2000 (24.08.2000)	Date of mailing of the international search report 01 September 2000 (01.09.2000)	
Name and mailing address of the ISA/ S.P.T.O	Authorized officer Telephone No.	

Form PCT/ISA/210 (second sheet) (July 1998)

INTERNATIONAL SEARCH REPORT

International application No. PCT/ES 99/00197
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Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: 1-9
because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- The additional search fees were accompanied by the applicant's protest.
 No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT Information on patent family members		International Application No PCT/ ES 00/00197	
Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9953944 A1	28.10.1999	AU 3148699 A ES 2138561 A EP 1002542 A	08.11.1999 01.01.2000 24.05.2000

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